

Evaluation of Chromium Induced Stress on Growth of Barley (*Hordeum vulgare* L.)

¹Anuradha, ²Ashish Tejasvi*

Author's Affiliation

^{1,2}Department of Botany, Agra College, Agra,
Uttar Pradesh 282001, India

*Corresponding Author:

Ashish Tejasvi

Department of Botany, Agra College,
Agra, Uttar Pradesh 282001, India

E-mail:

tejasvi.botany@gmail.com

Received on 15.08.2022

Revised on 10.11.2022

Accepted on 30.11.2022

Published on 15.12.2022

Keywords:

Chromium,
Fresh weight,
Heavy metal,
Hordeum vulgare L.,
Reduction.

Abstract

Heavy metals are harmful for plants growth. Chromium is one of them which affect growth and yield of plants negatively. Aim of present study was to access the effects of chromium on barley growth. Barley seeds were sown in pots filled with acid washed sand. Different levels (0, 25, 50, 75 and 100 μ M) of chromium treatments were applied. Chromium (III) chloride hexahydrate was taken as a source of chromium. Distilled water was used as control (0 μ M) treatment. Results showed that root, shoot and total plant length gets reduced significantly and the reduction percentage over control was also increased with increasing Cr concentrations. Leaf surface area was also decreased non-significantly. Chromium also caused a negative impact on fresh and dry weight of (root, shoot and whole plant). Root, shoot fresh weight was diminished by chromium non-significantly at lower concentrations (25 and 50 μ M) but significant reduction was observed at higher concentrations (75 and 100 μ M). Fresh weight of whole plant was reduced significantly over control by all chromium treatments (25, 50, 75 and 100 μ M). A significant reduction was also recorded for plant dry weight (root, shoot and whole plant).

How to cite this article: Anuradha and Tejasvi A. (2022). Evaluation of Chromium Induced Stress on Growth of Barley (*Hordeum vulgare* L.). *Bulletin of Pure and Applied Sciences-Botany*, 41B (2), 148-153.

INTRODUCTION

Various types of pollutants affect the environment; heavy metal emission due to urbanization is one of the major factors which cause serious problems for motherland and atmosphere (Kumar et al., 2015; Kumar et al., 2021b). Chromium has second rank in heavy metals which contaminated the soil, water through sedimentation (Kumar et al., 2017; Kumar et al., 2021a). Chromium is used in

various industries viz. leather, textile and steel industries. Overuse of fertilizers and pesticides created severe conditions (Nath et al., 2005; Shrestha et al., 2007; Ogundiran and Afolabi, 2008; Wang and Chen, 2009). Extreme amount of chromium reduced seed germination, plant growth and photosynthetic rate of plants. Chlorophyll content and enzymatic activities were also affected by chromium stress (Ali et al., 2012; Dotaniya et al., 2014; Ali et al., 2015). Phytotoxicity induced by chromium may cause

high oxidative injury in plant cells (Panda and Patra, 2002; Ali et al., 2011).

Chromium contaminated soil and water damage the crop tissues and very much harmful for human beings through food chains. When chromium absorbed by roots only a little amount of chromium translocate in higher parts of plants and great amount of chromium retained in roots. Chromium can be taken by plants in both Cr (III) and Cr (VI) form but there is no specific mechanism found in plants for chromium uptake (Vernay et al., 2007; Babula et al., 2008). The natural substrates can play significant role in the removal of different pollutants (Singh et al., 2018; Singh et al., 2021; Singh et al., 2022).

Barley belongs to grass family Poaceae and an important grain crop. It has fourth rank in production all around the world after maize, wheat and rice (Kumar et al., 2012). It has high nutritional value. It is also used for animal feeding, brewing, baking, confectionary, beverage, dairy products and condiments. Barley has a rich amount of soluble dietary fibers (Idehen et al., 2017; Tobiasz- Salach and Augustynska- Prejsnar, 2020). Object of the present investigation was to identify the chromium induced effects on barley growth.

MATERIAL AND METHODS

Seed of barley (*Hordeum vulgare* var. RD 2552) were collected from local market of Agra, Uttar Pradesh, India. Different seedling growth parameters were observed under different concentrations of chromium. To observe heavy metal tolerance of barley test cultivar, Chromium (III) chloride hexahydrate ($\text{CrCl}_3 \cdot 6\text{H}_2\text{O}$) was used as a source of chromium.

Experiment and Treatments

Experiments were conducted at Department of Botany, Agra College Agra, U.P., India. At first, seeds of barley (*Hordeum vulgare* var. RD 2552) were disinfected with mercuric chloride, and then washed with water to remove mercuric chloride. After that seeds were sowed in pots filled acid washed sand. The nutrients were provided by Long Aston nutrient solution

having KNO_3 , 5 mM; $\text{Ca}(\text{NO}_3)_2$, 4 mM; $\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$, 1.33mM; $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 3mM; Ferric citrate. $5\text{H}_2\text{O}$, 0.10 mM; MnSO_4 , 0.01; $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$, 0.001; $\text{Zn SO}_4 \cdot 7\text{H}_2\text{O}$, 0.001; $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24} \cdot 4\text{H}_2\text{O}$, 0.0002 mM and NaCl, 0.10 mM. Four treatments of Cr (25, 50, 75 and 100 μM) were given to evaluate the seedling characters of barley. Distilled water was used as control (0 μM) treatment. Three replicates were used for each treatment. The plants were taken out from sand after 21 DAS and different parameters under consideration were recorded.

Growth Parameters

Randomly, three plants were taken from each treatment to calculate the arithmetic mean for a respective parameter. Growth parameters recorded for study were as follows:

Root, shoot and total length of plant; Root, shoot and whole plant fresh weight; Root, shoot and whole plant dry weight and leaf surface area.

Statistical Analysis

The results of phytotoxicity under the chromium induced stress are shown as the mean \pm standard error (SE) with three replicates of each treatment and statistical analysis of collected data were exposed by using analysis of variances (ANOVA) and variances were determined by least significance differences (LSD) at 0.05 probability by using SPSS software.

RESULTS AND DISCUSSION

The impact of heavy metal (Cr) was observed on barley (*Hordeum vulgare* var. RD 2552) and its consequences were assessed on different plant growth parameters.

Plant Length and Leaf Surface Area

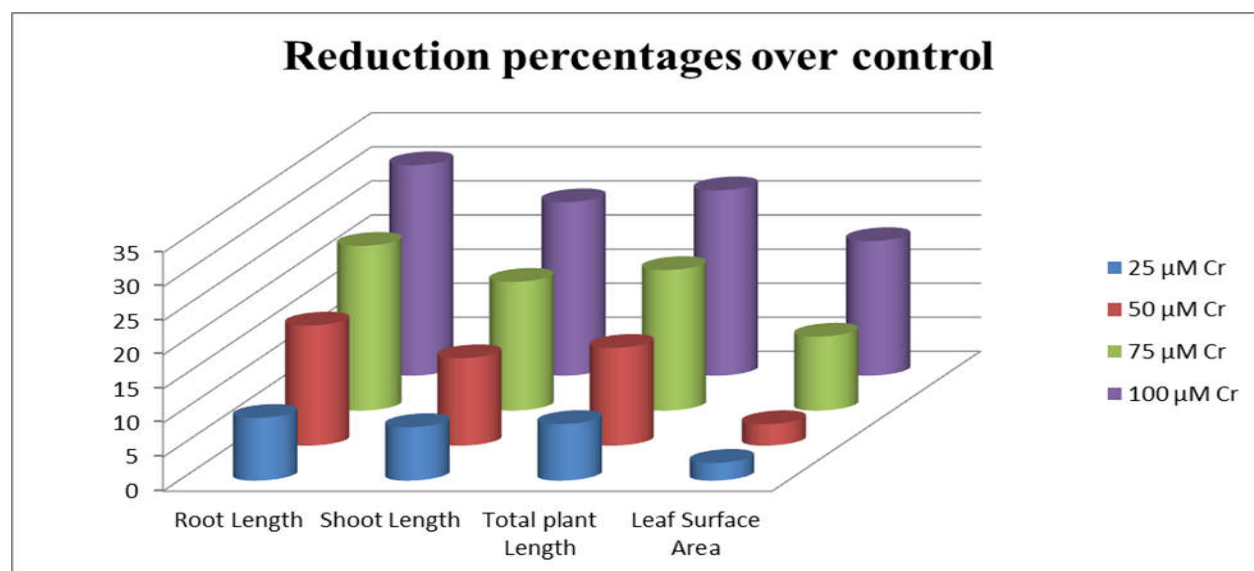
Root, shoot and leaves of barley are heavily affected by chromium stress (Table 1.). Root, shoot and total plant length; Leaf surface area were negatively influenced with increasing Cr concentrations. A significant retardation in plant length and leaf surface area was observed in all treatments (Figure 1.). Similar findings were observed earlier by Chatterjee and Chatterjee, 2000; Sundaramoorthy et al. 2010; Ding et al., 2019 and Singh et al., 2020.

Table 1: Effect of different concentrations of chromium on root length, shoot length, total plant length (cm) and leaf surface area (cm²) of barley

Chromium concentrations (μM)	Root length (cm)	Shoot length (cm)	Total length of plant (cm)	Leaf surface Area (cm ²)
Control (0)	8.97 ± 0.10	18.65 ± 0.17	27.62 ± 0.24	11.02 ± 0.72
25	8.14 ± 0.29*	17.18 ± 0.24*	25.32 ± 0.27*	10.73 ± 0.52 ^{ns}
50	7.39 ± 0.25*	16.25 ± 0.26*	23.65 ± 0.33*	10.67 ± 0.68 ^{ns}
75	6.80 ± 0.16*	15.11 ± 0.10*	18.54 ± 0.22*	9.82 ± 0.54 ^{ns}
100	6.20 ± 0.18*	13.90 ± 0.42*	17.01 ± 0.45*	8.84 ± 0.53 ^{ns}

Data is average of three replicates. Average ± Standard Error (SE)

* Significant at P < 0.05

**Figure 1:** Reduction percentages over control for root length, shoot length, total plant length and leaf surface area of barley

Fresh and dry weight

Fresh weight of root, shoot and whole plant of barley were decreased in all heavy metals stress treatments (Table 2. , Figure 2.). Fresh weight inhibition for 25 and 50 μM treatments was non-significant but significant at 75 and 100 μM concentrations. Dry weight of root, shoot and whole plant of barley followed the same pattern as for fresh weight resulted, the decrement in

dry weight of root, shoot and whole plant for all Cr concentrations. Retardation in dry weight of whole plant was non-significant at 25 μM but significant results were observed for 50, 75 and 100 μM. These results are in conformity with those of Vernay et al., 2007; Salvatore et al., 2008; Nematshahi et al., 2012; Gill et al., 2015; Ghani et al., 2017; Ding et al., 2019.

Table 2: Effect of different concentrations of chromium on fresh and dry weight of root, shoot and whole plant

Chromium concentrations (μM)	Fresh weight of root (g)	Fresh weight of shoot (g)	Total fresh weight/plant (g)	Dry weight of root (g)	Dry weight of shoot (g)	Total dry weight/plant (g)
Control (0)	0.086 \pm 0.0033	1.503 \pm 0.0232	1.59 \pm 0.0244	0.0128 \pm 0.0002	0.172 \pm 0.002	0.185 \pm 0.002
25	0.076 \pm 0.0026 ^{ns}	1.424 \pm 0.0195 ^{ns}	1.5 \pm 0.0197*	0.0119 \pm 0.0002*	0.162 \pm 0.0023*	0.177 \pm 0.0023 ^{ns}
50	0.067 \pm 0.0029 ^{ns}	1.29 \pm 0.0412*	1.36 \pm 0.0435*	0.0115 \pm 0.0002*	0.146 \pm 0.0035*	0.162 \pm 0.0033*
75	0.063 \pm 0.0022*	1.09 \pm 0.0389*	1.16 \pm 0.0394*	0.011 \pm 0.0002*	0.136 \pm 0.0029*	0.150 \pm 0.0029*
100	0.061 \pm 0.0022*	0.98 \pm 0.0279*	1.04 \pm 0.0264*	0.0103 \pm 0.0002*	0.122 \pm 0.0031*	0.134 \pm 0.0032*

Data is average of three replicates. Average \pm Standard Error (SE)

*Significant at $P < 0.05$

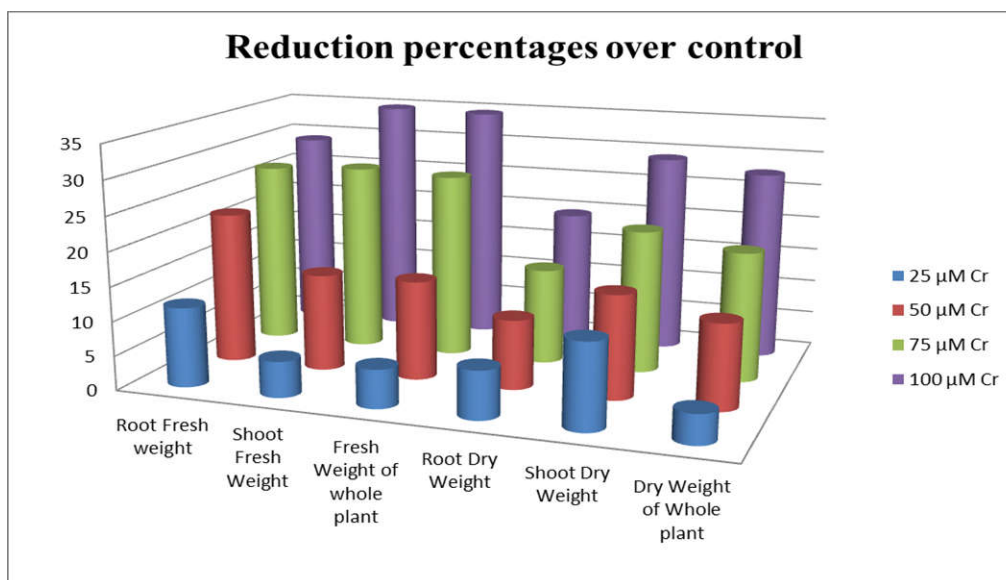


Figure 2: Reduction percentages over control for the fresh and dry weight of root, shoot and whole plant of barley

CONCLUSION

Heavy metals are toxic for plants. Chromium induced stressful conditions resulted in retardation of growth attributes of the test crop. Available literature of earlier studies concludes that heavy metals stress declined the crop productivity by producing reactive oxygen species (ROS). Present study is a short step which advocates the development of heavy metal resistant crop varieties.

REFERENCES

1. Ali, S., Cai, S., Zeng, F., Qiu, B. and Zhang, G.P. (2012). The effect of salinity and hexavalent chromium stresses on uptake and accumulation of mineral elements in barley genotypes differing in salt tolerance. *Journal Plant Nutrition*, 35, 827–839.
2. Ali, S., Chaudhary, A., Rizwan, M., Anwar, H.T., Adrees, M., Farid, M., Irshad, M.K.,

- Hayat, T. and Anjum, S.A. (2015). Alleviation of chromium toxicity by glycinebetaine is related to elevated antioxidant enzymes and suppressed chromium uptake and oxidative stress in wheat (*Triticum aestivum* L.). *Environmental Science and Pollution Research*, 22(14), 10669–10678. <https://doi.org/10.1007/s11356-015-4193-4>
3. Ali, S., Zeng, F., Cai, S., Qiu, B. and Zhang, G. (2011). The interaction of salinity and chromium in the influence of barley growth and oxidative stress. *Plant Soil Environment*, 57, 153–159.
4. Babula, P., Adam, V., Opatrilova, R., Zehnalek, J., Havel, L. and Kizek, R. (2008). Uncommon heavy metals, metalloids and their plant toxicity: A review. *Environmental Chemistry Letters*, 6, 189–213.
5. Chatterjee, J. and Chatterjee, C. (2000). Phytotoxicity of cobalt, chromium and copper in cauliflower. *Environmental Pollution*, 109, 69–74.
6. Ding, G., Jin, Z., Han, Y., Sun, P., Li, G. and Li, W. (2019). Mitigation of chromium toxicity in *Arabidopsis thaliana* by sulphur supplementation. *Ecotoxicology and Environmental Safety*, 182, 1–6.
7. Dotaniya, M.L., Das, H. and Meena, V.D. (2014). Assessment of chromium efficacy on germination, root elongation and coleoptile growth of wheat (*Triticum aestivum* L.) at different growth periods. *Environmental Monitoring and Assessment*, 186(5), 2957–2963. <https://doi.org/10.1007/s10661-013-3593-5>
8. Ghani, A., Hussain, M., Ikram, M., Yaqoob, M., Shaukat, R., Munawar, A., Ullah, R. and Imtiaz, A. (2017). Effect of chromium toxicity on the growth and mineral composition of brown mustard (*Brassica juncea* L.). *World Wide Journal of Multidisciplinary Research and Development*, 3(10), 36–38.
9. Gill, R.A., Zang, L., Ali, B., Farooq, M.A., Cui, P., Yang, S., Ali, S. and Zhou, W. (2015). Chromium-induced physio-chemical and ultrastructural changes in four cultivars of *Brassica napus* L. *Chemosphere*, 120, 154–164. <https://doi.org/10.1016/j.chemosphere.2014.06.029>
10. Idehen, E., Tang, Y. and Sang, S. (2017). Bioactive phytochemicals in barley. *Journal of Food Drug Analysis*, 25(1), 148–161.
11. Kumar, A., Singh, R., Upadhyay, S. K., Kumar S. and Charaya, M.U. (2021b). Biosorption: The removal of toxic dyes from industrial effluent using phyto-biomass- A Review. *Plant Archives*, 21(1), 1320–132.
12. Kumar, M., Mukherjee, T.K., Sharma, I., Upadhyay, S.K. and Singh, R. (2021a). Role of bacteria in bioremediation of chromium from wastewaters: An overview. *Bio-Science Research Bulletin*, 37(2), 77–87.
13. Kumar, P., Bhati, H., Rani A. and Singh, R. (2015). Role of Biosorption of dyes and microorganisms in environment. *Advances in Life Sciences*, 4(2), 38–41.
14. Kumar, P., Malik, M., Singh, R., Rani A. and Kumar, A. (2017). A comparative study on the biosurfactant producing bacteria from oil contaminated water. *Bio- Science Research Bulletin*. 33 (1), 37–43.
15. Kumar, S., Mishra, C.N., Sarkar, B. and Singh, S.S. (2012). Barley (*Hordeum vulgare* L.). *Breeding Indian Field crops - (Hordeum vulgare L.)*. Chapter, 1–18.
16. Nath, K., Saini, S. and Sharma, Y.K. (2005). Chromium in tannery industry effluent and its effect on plant metabolism and growth. *Journal of Environmental Biology*, 26(2), 197–204.
17. Nematshahi, N., Lahouti, M., Ganjeali, A. (2012). Accumulation of chromium and its effect on growth of (*Allium cepa* cv. Hybrid), *European Journal of Experimental Biology*, 2(4), 969–974.
18. Ogundiran, O.O. and Afolabi, T.A. (2008). Assessment of the physicochemical parameters and heavy metals toxicity of leachates from municipal solid waste open dumpsite. *International Journal of Environmental Science & Technology*, 5, 243–250.
19. Panda, S.K. and Patra, H.K. (2002). Does Cr (III) produce oxidative damage in excised wheat leaves? *Journal of Plant Biology*, 27, 105–110.
20. Salvatore, M.D., Carafa, A.M. and Carratu, G. (2008). Assessment of heavy metals phytotoxicity using seed germination and root elongation tests: a comparison of two

- growth substrates. *Chemosphere*, 73(9), 1461-1464.
21. Shrestha, R., Fischer, R. and Sillanpaa, M. (2007). Investigations on different positions of electrodes and their effects on the distribution of Cr at the water sediment interface. *International Journal of Environmental Science Technology*, 4, 413-420.
 22. Singh, D., Sharma, N.L., Singh, C.K., Sarkar, S.K., Singh, I. and Dotaniya, M.L. (2020). Effect chromium (VI) toxicity on morpho-physiological characteristics, yield, and yield components of two chickpea (*Cicer arietinum* L.) varieties. *PLoSOne*, 15(12), 1-27. e0243032. <https://doi.org/10.1371/journal.pone.0243032>
 23. Singh, M., Kumari, M., Upadhyay, S. K., Yadav, M. and Singh, R. (2022). Biosorption of malachite green using marine macroalgae from aqueous solution. *International Journal of Botany Studies*, 7(2):402-405.
 24. Singh, R., Upadhyay, S. K., Rani, A., Kumar, P., Kumar, A Singh, C. (2018). Lignin biodegradation in nature and significance. *Vegetos*, 31(4), 39-44.
 25. Singh, R., Upadhyay, S. K., Singh, M., Sharma, I., Sharma, P., Kamboj, P., Saini, A., Voraha, R., Sharma, A.K., Upadhyay, A.K. and Khan, F. (2021). Chitin, chitinases and chitin derivatives in biopharmaceutical, agricultural and environmental perspective. *Biointerface Research in Applied Chemistry*, 11(3), 9985-10005.
 26. Sundaramoorthy, P., Chidambaram, A., Ganesh, K.S., Unnikannan, P. and Baskaran, L. (2010). Chromium stress in paddy: (i) Nutrient status of paddy under chromium stress; (ii) Phytoremediation of chromium by aquatic and terrestrial weeds. *Comptes Rendus Biologies*, 333 (8), 597-607.
 27. Tobiasz-Salach, R. and Augustynska-Prejsnar, A. (2020). Response of spring barley to foliar fertilization with Cu and Mn. *Acta Sci. Pol. Agricultura*, 19(1), 29-39.
 28. Vernay, P., Gauthier-Moussard, C. and Hitmi, A. (2007). Interaction of bioaccumulation of heavy metal chromium with water relation, mineral nutrition and photosynthesis in developed leaves of *Lolium perenne* L. *Chemosphere*, 68, 1563-1575.
 29. Wang, J. and Chen, C. (2009). Biosorbents for heavy metals removal and their future. *Biotechnology Advances*, 27(2), 195-226. <https://doi.org/10.1016/j.biotechadv.2008.11.002>
